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Metabotropic glutamate receptors: intracellular signaling pathways

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Metabotropic glutamate receptors are classified into three groups, primarily on the basis of sequence similarity and whether they positively couple to the phospholipase C cascade or negatively couple to adenylyl cyclases. The past decade of research, drawing on sophisticated molecular approaches, has revealed a multitude of additional intracellular components that assemble as protein scaffolds around neuronal metabotropic glutamate receptors, establishing functional links to postsynaptic density structures, to membrane-bound enzymes and ion channels, and to the nucleus. Characterization of these novel transduction mechanisms is providing new insights into the roles of metabotropic glutamate receptors in the regulation and modulation of diverse functions in the nervous system.

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Introduction

The metabotropic glutamate receptors (mGluRs) were discovered when it was observed that exposing neurons to glutamate activated not only ionotropic receptors but also stimulated phospholipase C (PLC) [1,2]. Soon thereafter, a family of eight distinct mGluR subtypes was identified, and the palette of associated intracellular signaling mechanisms was greatly extended [3]. The mGluRs are classified according to structural and functional criteria into Group I (mGluR1 and mGluR5), Group II (mGluR2 and mGluR3), and Group III (mGluR4, mGluR6, mGluR7, and mGluR8) [4]. Here, we provide a brief update of new developments that expand and clarify our understanding of the transduction mechanisms mediating responses initiated by mGluRs. A major advance in this field, the retrograde signaling by endocannabinoids following the activation of postsynaptic

mGluRs, will not be discussed explicitly here as this topic is the subject of several comprehensive recent reviews (e.g. see [5]).

Transduction within the mGluR

Upon binding of glutamate, a conformational change in homodimeric mGluRs promotes the coupling of G proteins to specific intracellular domains. Structural studies, beginning with the crystallization and characterization of the agonist-bound and ‘unliganded’ forms of the glutamate binding site of mGluR1, provided initial insights into the underlying process [6]. Agonist binding stabilizes the closed conformation of the extracellular domain and results in G protein activation that is dependent upon a disulfide bridge between conserved cysteine residues in the extracellular agonist binding loop and the third transmembrane domain [6,7]. This disulfide bridge mediates intrareceptor signaling by inducing an allosteric interaction between the glutamate binding domain and the heptahelical domain [7]. Thus, agonist binding changes the relative positions of the helical domains of these dimeric receptors to permit G protein activation.

A peculiar property reported for several metabotropic responses, including those mediated by mGluRs [8], is their voltage sensitivity. It has now been shown that this voltage dependence resides within the receptor itself. It appears that depolarization modifies the conformation of the second and third intracellular loops, thus affecting the association with G proteins [9]. These depolarization-dependent changes in G protein binding, in turn, alter the proportion of receptors in the high-affinity state for agonist [10].

G-protein-independent signaling

The canonical cascade coupling metabotropic receptors with their intracellular effectors begins with the activation of G proteins, hence the name G-protein-coupled receptors. Over the past decade, however, several studies have reported metabotropic responses that do not involve G proteins [11]. Evidence that mGluRs can also function in this manner came from experiments showing that activation of mGluR1 in hippocampal neurons simultaneously triggers both G-protein-dependent and -independent signaling to induce distinct currents [12,13]. The same conclusion was reached in a study using hippocampal pyramidal neurons from transgenic mice lacking the G proteins associated with postsynaptic mGluRs, in which inward currents mediated by mGluRs nevertheless persisted [14]. It is interesting that, for certain neuronal responses, specific mGluRs appear to

preferentially utilize G-protein-dependent pathways whereas, for other responses, the G-protein-independent mechanism predominates. For example, activation of mGluRs can lead to potentiation of responses mediated by N-methyl-D-aspartate (NMDA)-type glutamate receptors, which are critical for the induction of many forms of synaptic plasticity. In CA3 pyramidal neurons, which express both mGluR1 and mGluR5, NMDA receptor potentiation by mGluR5 was found to be G-protein-dependent, whereas potentiation by mGluR1 could proceed independently of G protein activation [15]. In the same cells, a mGluR1- and mGluR5-induced cationic current necessitates the cooperative activation of both G-protein-independent and G-protein-dependent pathways, with the former targeting calcium-sensitive cationic channels that conduct the current and the latter eliciting the release of the requisite calcium from intracellular stores (Figure 1) [16].

β -arrestins and mGluR signaling

Following their activation, metabotropic receptors undergo rapid desensitization through a process involving phosphorylation by G-protein-coupled receptor kinases,

which then allows the binding of the adaptor proteins β -arrestin 1 and β -arrestin 2 that direct receptor endocytosis through targeting to clathrin-coated pits [17]. This mechanism also holds for the mGluRs, although a phosphorylation-independent form of desensitization mediated by G-protein-coupled receptor kinase-2 has been observed, as recently reviewed [18]. In addition to their role in receptor desensitization, β -arrestins act as scaffolding elements for the recruitment of signaling proteins that regulate diverse cellular functions [11,17,18]. A variety of direct and indirect evidence indicates that mGluRs associate with β -arrestins in the initiation of intracellular cascades effecting neuronal responses. Both the G-protein-independent actions of mGluR1 that lead to the induction of cationic current [12] and the potentiation of NMDA currents [15] in the hippocampus were shown to require activation of the non-receptor tyrosine kinase Src. In this case, it is likely that β -arrestin is acting as the adaptor to couple a Src-family kinase to the activated mGluR, as has been shown for Src activation by numerous other metabotropic receptors [11,17]. A more recent study has identified a role for β -arrestin 2 in the recruitment of Src to Group III mGluRs, leading to the activation of mitogen-activated protein kinase (MAPK) pathways [19].

Figure 1

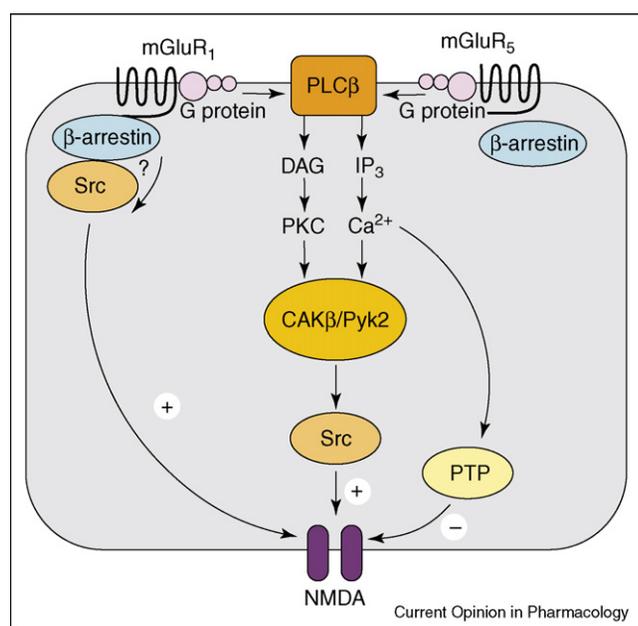


Diagram illustrating the mechanisms proposed to underlie the modulation of NMDA responses by Group I mGluRs. The G-protein-dependent activation of PLC β triggers divergent signaling cascades leading both to potentiation of NMDA responses (centre of scheme) and to calcium-dependent activation of tyrosine phosphatases that depress NMDA responses. When G proteins are experimentally blocked, a parallel pathway utilized by mGluR1 is revealed (left-hand side of scheme) that activates Src, probably via the adaptor protein β -arrestin, leading to potentiation of NMDA responses. CAK β /Pyk2, cell adhesion kinase β /proline-rich tyrosine kinase; DAG, diacylglycerol; PKC, protein kinase C; PTP, protein tyrosine phosphatase. Note that, for simplicity, mGluRs which function as dimers are depicted as monomers.

Ubiquitous actions of calcium

Almost every step in the signaling pathways associated with mGluRs requires, or is modulated by, calcium. Beginning with the receptors themselves, both the potency and efficacy of glutamate action at Group I mGluRs is enhanced with increasing concentrations of extracellular calcium [20]. Importantly, the efficacy of mGluR signaling is modulated by physiologically relevant changes in extracellular calcium, such that calcium depletion in the synaptic cleft, as occurs during burst firing, causes significant inhibition of postsynaptic mGluR function [21]. Intracellularly, significant release of calcium is observed after synaptic activation of dendritic mGluRs, which can propagate as waves and even reach the cell nucleus under appropriate conditions [22].

Recent studies have shed light on the modulation by calcium of the transduction pathway between mGluRs and NMDA receptors. Work performed by John MacDonald and colleagues has delineated a transduction pathway that potentiates NMDA receptor currents through the sequential activation of metabotropic receptors, PLC β , protein kinase C, CAK β /Pyk2 (cell adhesion kinase β /proline-rich tyrosine kinase) and Src [23,24]. Interestingly, when initiated by mGluRs, this pathway requires inositol-1,4,5-trisphosphate (IP₃) receptor-dependent intracellular calcium release; calcium influx through NMDA receptors or voltage-dependent calcium channels does not lead to potentiation of NMDA responses [25]. However, if intracellular calcium rises excessively, an antagonistic G-protein-dependent pathway prevails that reduces NMDA responses [15,26].

Thus, neurons that express NMDA receptors contain both mGluR-dependent facilitatory and depressing pathways to ensure the precise regulation of this physiologically crucial receptor. Furthermore, either the facilitating or the depressing pathway can dominate depending upon the cell type [26], and is likely to be regulated by factors such as differences in calcium signaling pathways or in intrinsic calcium buffering capacity. Such bidirectional modulation under the control of intracellular calcium concentration was also found to determine whether mGluR5-dependent plasticity of NMDA responses results in long-term potentiation (LTP) or long-term depression (LTD) at the perforant path–granule cell synapse [27]. These findings provide an explanation for the discrepancies in the literature concerning facilitatory versus depressing NMDA receptor modulation by mGluRs, and highlight the significance of differences in preparations and experimental conditions that could influence ambient intracellular calcium concentrations. Differences in NMDA receptor modulation could also contribute to the distinct susceptibilities of various neuronal cell types to ischemic cell death. Indeed, in hippocampal CA1 pyramidal neurons, transient energy deprivation results in Src-dependent upregulation of NMDA receptor function associated with delayed neuronal death whereas, in neighboring CA3 pyramidal neurons, which are known to be more resistant to ischemia, the same protocol activates tyrosine phosphatases and does not lead to either NMDA receptor potentiation or cell death [28]. The calcium-sensitive equilibrium between tyrosine kinases and phosphatases might also be important in determining the phosphorylation state of α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors, which in turn will determine whether synapses are in a potentiated [29] or depressed state [30].

Modulation of transcription and translation by mGluRs

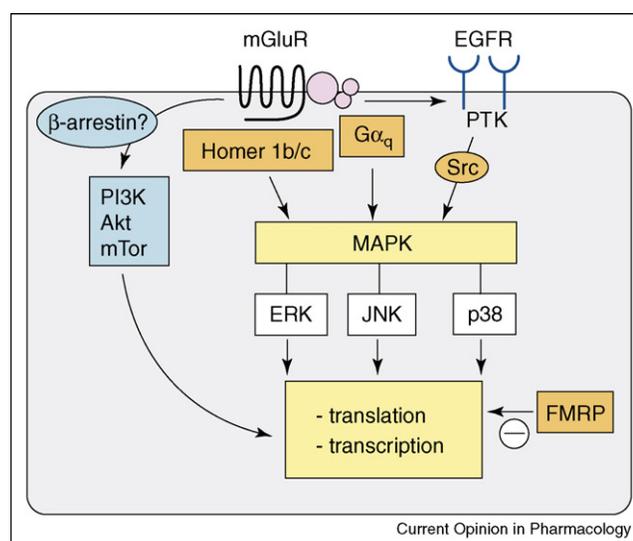
The consolidation of synaptic plasticity is dependent upon protein synthesis. The ability of mGluRs to initiate this process was shown by studies demonstrating a role for mGluR-dependent protein synthesis in the maintenance of epileptiform discharge [31] and in hippocampal synaptic plasticity [32,33]. Interestingly, the protein synthesis necessary for mGluR-dependent hippocampal LTD depends upon local translation of mRNA near the synapse, and not upon transcription [32]. Concurrently, it was reported that mGluR-dependent LTP was reduced after blockade of extracellular signal-regulated kinase (ERK)1/2–MAPK signaling [34]. More recent studies have refined our understanding of the signaling mechanisms linking Group I mGluRs to MAPK activation and the role that these pathways play in protein synthesis-dependent neuronal plasticity. MAPK cascades are triggered by stimuli at the extracellular membrane and culminate in the phosphorylation and activation of MAPKs comprising ERKs, c-Jun N-terminal kinases (JNKs) and p38s, which promote

translation and activate transcription factors to increase protein synthesis. Stimulation of mGluR5 leads to weak activation of ERK1/2 through the PLC β /IP $_3$ /Ca $^{2+}$ pathway and much stronger activation via the scaffolding protein Homer 1b/c [35]. Conversely, Homer 1a inhibits mGluR-dependent activation of MAPK, a mechanism important in downregulating chronic pain signaling [36]. Robust activation of ERK and JNK MAPKs is also achieved by mGluR5-dependent transactivation of the epidermal growth factor receptor [37,38].

The mGluR-dependent synaptic plasticity associated with persistent epileptiform discharge [39], hippocampal LTD [40] and hippocampal LTP in oriens/alveus interneurons [29] has been shown to depend both upon ERK activation and upon a transduction pathway that employs a tyrosine kinase, rather than PKC, to phosphorylate and activate ERK1/2. In addition, Group I mGluRs activate a transduction pathway involving phosphoinositide 3-kinase (PI3K), Akt and mammalian target of rapamycin (mTor), which modulates mRNA translation in parallel with the ERK pathway to induce LTD [41]. Several studies have also shown the involvement of p38 MAPKs in mGluR-dependent LTD (Figure 2) [42–44,45].

Translation and transcription factors targeted by MAPK cascades following mGluR activation have recently been

Figure 2



Activation of mGluRs can modulate both local postsynaptic translation of mRNA and nuclear transcription. The underlying signaling pathways typically target MAPKs, either originating at mGluRs via Homer or G protein activation, or following transactivation of the epidermal growth factor receptor (EGFR), thus inducing protein tyrosine kinase (PTK) activity of the intracellular domain and leading to Src activation. The fragile X mental retardation protein (FMRP) modulates mGluR signaling by virtue of its inhibitory effects on translation. The PI3K–Akt–mTor pathway represents a further signaling cascade that regulates translation in parallel to the MAPK pathways.

characterized. Activity of the cap-dependent translation protein eIF4E was shown to be under the control of both the ERK pathway [32,46^{*}] and the PI3K–Akt–mTor pathway [46^{*}], a finding which clarifies the mechanism underlying mGluR-dependent LTD. The mGluR-dependent phosphorylation of JNK increases transcription mediated by activator protein-1 [38], and activation of p38 regulates nuclear factor- κ B (NF- κ B) [45^{**}]. In knockout mice lacking the NF- κ B member c-Rel, late-phase LTD, as well as performance in a passive avoidance task, was diminished [45^{**}]. An additional factor modulating mGluR-dependent protein synthesis is a negative regulator of translation called fragile X mental retardation protein (FMRP), the levels of which rise in dendrites following activation of group I mGluRs. Thus, in mice lacking FMRP, mGluR-dependent LTD is enhanced [47], as is the propensity for epileptiform activity owing to over-activation of protein translation [48^{*}]. These findings suggest that antagonists of either Group I mGluRs or the downstream elements of the ERK signaling pathway may be useful in the treatment of fragile X syndrome [49].

mGluRs can also target transcription factors independent of MAPK or PI3K activation. Enhanced activation of mGluR4 in developing cerebellar granule cell cultures reduces Gli-1, a transcription factor in the Sonic Hedge Hog pathway [50]. This mGluR4-dependent effect was associated with reduced proliferation of cerebellar neural precursor cells and an increase in their differentiation into mature granule cells [50].

Conclusions and outlook

mGluRs play key roles in the modulation of diverse cellular responses. Recent structural advances have provided new insights into how changes in receptor conformation can initiate response transduction. In addition, molecular analysis is providing rich detail into the surprisingly divergent signaling pathways employed by these receptors, which modulate targets not only in the membrane but also in the cytoplasm and nucleus.

Although metabotropic receptors and the responses they mediate are becoming well characterized, the conditions under which mGluRs are synaptically activated remain to be established. Postsynaptic mGluRs are located at perisynaptic or extrasynaptic sites and, as such, will sense relatively low concentrations of glutamate diffusing out of the synaptic cleft. Yet, in most studies, high concentrations of agonist are applied to preparations that often exceed the EC₅₀ values for mGluRs by one to two orders of magnitude [4]. A challenge for future studies will be to design experiments that mimic mGluR activation levels occurring during physiological network activity.

An important focus of current research is the linking of molecular data on mGluRs with specific sensory

and behavioral functions. Apart from enhancing our understanding of diverse neuronal mechanisms, this approach is generating leads that promise new therapies for the treatment of a wide spectrum of psychiatric and neurological disorders [51].

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References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Sladeczek F, Pin JP, Recasens M, Bockaert J, Weiss S: **Glutamate stimulates inositol phosphate formation in striatal neurons.** *Nature* 1985, **317**:717-719.
2. Nicoletti F, Iadarola MJ, Wroblewski JT, Costa E: **Excitatory amino acid recognition sites coupled with inositol phospholipid metabolism: developmental changes and interaction with alpha 1-adrenoceptors.** *Proc Natl Acad Sci USA* 1986, **83**:1931-1935.
3. Nakanishi S: **Metabotropic glutamate receptors: synaptic transmission, modulation, and plasticity.** *Neuron* 1994, **13**:1031-1037.
4. Conn PJ, Pin JP: **Pharmacology and functions of metabotropic glutamate receptors.** *Annu Rev Pharmacol Toxicol* 1997, **37**:205-237.
5. Alger BE: **Retrograde signaling in the regulation of synaptic transmission: focus on endocannabinoids.** *Prog Neurobiol* 2002, **68**:247-286.
6. Kunishima N, Shimada Y, Tsuji Y, Sato T, Yamamoto M, Kumasaka T, Nakanishi S, Jingami H, Morikawa K: **Structural basis of glutamate recognition by a dimeric metabotropic glutamate receptor.** *Nature* 2000, **407**:971-977.
7. Rondard P, Liu J, Huang S, Malhaire F, Vol C, Pinault A, Labesse G, Pin JP: **Coupling of agonist binding to effector domain activation in metabotropic glutamate-like receptors.** *J Biol Chem* 2006, **281**:24653-24661.
8. Perroy J, Richard S, Nargeot J, Bockaert J, Fagni L: **Permissive effect of voltage on mGlu 7 receptor subtype signaling in neurons.** *J Biol Chem* 2002, **277**:1223-1228.
9. Ohana L, Barchad O, Parnas I, Parnas H: **The metabotropic glutamate G-protein-coupled receptors mGluR3 and mGluR1a are voltage sensitive.** *J Biol Chem* 2006, **281**:24204-24215.
10. De Lean A, Stadel JM, Lefkowitz RJ: **A ternary complex model explains the agonist-specific binding properties of the adenylate cyclase-coupled beta-adrenergic receptor.** *J Biol Chem* 1980, **255**:7108-7117.
11. Hall RA, Premont RT, Lefkowitz RJ: **Heptahelical receptor signaling: beyond the G protein paradigm.** *J Cell Biol* 1999, **145**:927-932.
12. Heuss C, Scanziani M, Gähwiler BH, Gerber U: **G-protein-independent signaling mediated by metabotropic glutamate receptors.** *Nat Neurosci* 1999, **2**:1070-1077.
13. Gee CE, Lacaille JC: **Group I metabotropic glutamate receptor actions in oriens/alveus interneurons of rat hippocampal CA1 region.** *Brain Res* 2004, **1000**:92-101.
14. Krause M, Offermanns S, Stocker M, Pedarzani P: **Functional specificity of G alpha q and G alpha 11 in the cholinergic and glutamatergic modulation of potassium currents and excitability in hippocampal neurons.** *J Neurosci* 2002, **22**:666-673.

15. Benquet P, Gee CE, Gerber U: **Two distinct signaling pathways upregulate NMDA receptor responses via two distinct metabotropic glutamate receptor subtypes.** *J Neurosci* 2002, **22**:9679-9686.
16. Gee CE, Benquet P, Gerber U: **Group I metabotropic glutamate receptors activate a calcium-sensitive TRP-like conductance in rat hippocampus.** *J Physiol* 2003, **546**:655-664.
17. Lefkowitz RJ, Shenoy SK: **Transduction of receptor signals by β -arrestins.** *Science* 2005, **308**:512-517.
18. Dhami GK, Ferguson SS: **Regulation of metabotropic glutamate receptor signaling, desensitization and endocytosis.** *Pharmacol Ther* 2006, **111**:260-271.
19. Jiang Q, Yan Z, Feng J: **Activation of group III metabotropic glutamate receptors attenuates rotenone toxicity on dopaminergic neurons through a microtubule-dependent mechanism.** *J Neurosci* 2006, **26**:4318-4328.
20. Hermans E, Challiss RA: **Structural, signalling and regulatory properties of the group I metabotropic glutamate receptors: prototypic family C G-protein-coupled receptors.** *Biochem J* 2001, **359**:465-484.
21. Hardingham NR, Bannister NJ, Read JC, Fox KD, Hardingham GE, Jack JJ: **Extracellular calcium regulates postsynaptic efficacy through group I metabotropic glutamate receptors.** *J Neurosci* 2006, **26**:6337-6345.
- By recording from synaptically coupled cortical cell pairs, the authors show that physiological fluctuations in extracellular calcium modulate postsynaptic mGluR function, leading to changes in AMPA-receptor-mediated synaptic potentials.
22. Watanabe S, Hong M, Lasser-Ross N, Ross WN: **Modulation of calcium wave propagation in the dendrites and to the soma of rat hippocampal pyramidal neurons.** *J Physiol* 2006, **575**:455-468.
23. Lu WY, Xiong ZG, Lei S, Orser BA, Dudek E, Browning MD, MacDonald JF: **G-protein-coupled receptors act via protein kinase C and Src to regulate NMDA receptors.** *Nat Neurosci* 1999, **2**:331-338.
24. Kotecha SA, MacDonald JF: **Signaling molecules and receptor transduction cascades that regulate NMDA receptor-mediated synaptic transmission.** *Int Rev Neurobiol* 2003, **54**:51-106.
25. Guo W, Wei F, Zou S, Robbins MT, Sugiyono S, Ikeda T, Tu JC, Worley PF, Dubner R, Ren K: **Group I metabotropic glutamate receptor NMDA receptor coupling and signaling cascade mediate spinal dorsal horn NMDA receptor 2B tyrosine phosphorylation associated with inflammatory hyperalgesia.** *J Neurosci* 2004, **24**:9161-9173.
26. Grishin AA, Gee CE, Gerber U, Benquet P: **Differential calcium-dependent modulation of NMDA currents in CA1 and CA3 hippocampal pyramidal cells.** *J Neurosci* 2004, **24**:350-355.
27. Harney SC, Rowan M, Anwyl R: **Long-term depression of NMDA receptor-mediated synaptic transmission is dependent on activation of metabotropic glutamate receptors and is altered to long-term potentiation by low intracellular calcium buffering.** *J Neurosci* 2006, **26**:1128-1132.
28. Gee CE, Benquet P, Raineteau O, Rietschin L, Kirbach SW, Gerber U: **NMDA receptors and the differential ischemic vulnerability of hippocampal neurons.** *Eur J Neurosci* 2006, **23**:2595-2603.
29. Topolnik L, Azzi M, Morin F, Kougioumoutzakis A, Lacaille JC: **mGluR1/5 subtype-specific calcium signalling and induction of long-term potentiation in rat hippocampal oriens/alevis interneurons.** *J Physiol* 2006, **575**:115-131.
30. Moulton PR, Gladding CM, Sanderson TM, Fitzjohn SM, Bashir ZI, Molnar E, Collingridge GL: **Tyrosine phosphatases regulate AMPA receptor trafficking during metabotropic glutamate receptor-mediated long-term depression.** *J Neurosci* 2006, **26**:2544-2554.
31. Merlin LR, Bergold PJ, Wong RKS: **Requirement of protein synthesis for group I mGluR-mediated induction of epileptiform discharges.** *J Neurophysiol* 1998, **80**:989-993.
32. Huber KM, Kayser MS, Bear MF: **Role for rapid dendritic protein synthesis in hippocampal mGluR-dependent long-term depression.** *Science* 2000, **288**:1254-1257.
33. Raymond CR, Thompson VL, Tate WP, Abraham WC: **Metabotropic glutamate receptors trigger homosynaptic protein synthesis to prolong long-term potentiation.** *J Neurosci* 2000, **20**:969-976.
34. Coogan AN, O'Leary DM, O'Connor JJ: **P42/44 MAP kinase inhibitor PD98059 attenuates multiple forms of synaptic plasticity in rat dentate gyrus *in vitro*.** *J Neurophysiol* 1999, **81**:103-110.
35. Mao L, Yang L, Tang Q, Samdani S, Zhang G, Wang JQ: **The scaffold protein Homer1b/c links metabotropic glutamate receptor 5 to extracellular signal-regulated protein kinase cascades in neurons.** *J Neurosci* 2005, **25**:2741-2752.
36. Tappe A, Klugmann M, Luo C, Hirlinger D, Agarwal N, Benrath J, Ehrenguber MU, Doring MJ, Kuner R: **Synaptic scaffolding protein Homer1a protects against chronic inflammatory pain.** *Nat Med* 2006, **12**:677-681.
- This study shows that Homer 1a is rapidly upregulated in spinal neurons in response to peripheral inflammation, leading to reduced MAPK activation and decreased expression of dendritic spines. The reduction in synaptic contacts is consistent with a negative feedback loop to limit pain signaling.
37. Peavy RD, Chang MS, Sanders-Bush E, Conn PJ: **Metabotropic glutamate receptor 5-induced phosphorylation of extracellular signal-regulated kinase in astrocytes depends on transactivation of the epidermal growth factor receptor.** *J Neurosci* 2001, **21**:9619-9628.
38. Yang L, Mao L, Chen H, Catavsan M, Kozinn J, Arora A, Liu X, Wang JQ: **A signaling mechanism from G alpha q-protein-coupled metabotropic glutamate receptors to gene expression: role of the c-Jun N-terminal kinase pathway.** *J Neurosci* 2006, **26**:971-980.
39. Zhao W, Bianchi R, Wang M, Wong RKS: **Extracellular signal-regulated kinase 1/2 is required for the induction of group I metabotropic glutamate receptor-mediated epileptiform discharges.** *J Neurosci* 2004, **24**:76-84.
40. Gallagher SM, Daly CA, Bear MF, Huber KM: **Extracellular signal-regulated protein kinase activation is required for metabotropic glutamate receptor-dependent long-term depression in hippocampal area CA1.** *J Neurosci* 2004, **24**:4859-4864.
41. Hou L, Klann E: **Activation of the phosphoinositide 3-kinase-Akt-mammalian target of rapamycin signaling pathway is required for metabotropic glutamate receptor-dependent long-term depression.** *J Neurosci* 2004, **24**:6352-6361.
42. Bolshakov VY, Carboni L, Cobb MH, Siegelbaum SA, Belardetti F: **Dual MAP kinase pathways mediate opposing forms of long-term plasticity at CA3-CA1 synapses.** *Nat Neurosci* 2000, **3**:1107-1112.
43. Rush AM, Wu J, Rowan MJ, Anwyl R: **Group I metabotropic glutamate receptor (mGluR)-dependent long-term depression mediated via p38 mitogen-activated protein kinase is inhibited by previous high-frequency stimulation and activation of mGluRs and protein kinase C in the rat dentate gyrus *in vitro*.** *J Neurosci* 2002, **22**:6121-6128.
44. Huang CC, You JL, Wu MY, Hsu KS: **Rap1-induced p38 mitogen-activated protein kinase activation facilitates AMPA receptor trafficking via the GDI.Rab5 complex. Potential role in (S)-3,5-dihydroxyphenylglycine-induced long term depression.** *J Biol Chem* 2004, **279**:12286-12292.
45. O'Riordan KJ, Huang IC, Pizzi M, Spano P, Boroni F, Egli R, Desai P, Fitch O, Malone L, Ahn HJ *et al.*: **Regulation of nuclear factor kappaB in the hippocampus by group I metabotropic glutamate receptors.** *J Neurosci* 2006, **26**:4870-4879.
- This elegant study maps a pathway linking mGluR5 activation to stimulation of NF- κ B. This pathway is involved in the maintenance of LTD and its disruption worsens memory impairment in mice.
46. Banko JL, Hou L, Poulin F, Sonenberg N, Klann E: **Regulation of eukaryotic initiation factor 4E by converging signaling**

pathways during metabotropic glutamate receptor-dependent long-term depression. *J Neurosci* 2006, **26**:2167-2173.

It has been known for several years that mGluR activation can trigger protein synthesis in the vicinity of synapses. This study identifies cap-dependent translation as a key mechanism in this process.

47. Huber KM, Gallagher SM, Warren ST, Bear MF: **Altered synaptic plasticity in a mouse model of fragile X mental retardation.** *Proc Natl Acad Sci USA* 2002, **99**:7746-7750.
48. Chuang SC, Zhao W, Bauchwitz R, Yan Q, Bianchi R, Wong RK:
 - **Prolonged epileptiform discharges induced by altered group I metabotropic glutamate receptor-mediated synaptic responses in hippocampal slices of a fragile X mouse model.** *J Neurosci* 2005, **25**:8048-8055.
49. Bear MF, Huber KM, Warren ST: **The mGluR theory of fragile X mental retardation.** *Trends Neurosci* 2004, **27**:370-377.
50. Canudas AM, Di Giorgi-Gerevini V, Iacovelli L, Nano G, D'Onofrio M, Arcella A, Giangaspero F, Busceti C, Ricci-Vitiani L, Battaglia G *et al.*: **PHCC, a specific enhancer of type 4 metabotropic glutamate receptors, reduces proliferation and promotes differentiation of cerebellar granule cell neuroprecursors.** *J Neurosci* 2004, **24**:10343-10352.
51. Niswender CM, Jones CK, Conn PJ: **New therapeutic frontiers for metabotropic glutamate receptors.** *Curr Top Med Chem* 2005, **5**:847-857.

This paper provides further evidence for an interaction between mGluR signaling pathways and FMRP, in this case revealing a greater propensity for seizures in the absence of FMRP.